## Spongiacysteine, a Novel Cysteine Derivative from Marine Sponge Spongia sp.

Keiko Kobayashi, Hiroki Shimogawa, Akira Sakakura, Toshiaki Teruya, Kiyotake Suenaga, and Hideo Kigoshi\* Department of Chemistry, University of Tsukuba, Tennoudai, Tsukuba 305-8571

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A novel cysteine derivative, spongiacysteine (1), was isolated from marine sponge *Spongia* sp. The gross structure was elucidated by detailed spectroscopic analysis, and the absolute stereostructure was established by its total synthesis. This compound showed moderate antimicrobial activity against rice blast fungus *Pyricularia oryzae*.

Marine sponges are a prolific source of new compounds with diverse structures that often have interesting biological activities. In our continuing search for new substances from marine sponges,<sup>1</sup> we investigated the constituents of marine sponge *Spongia* sp.<sup>2</sup> collected at Tateyama beach, Chiba Prefecture, and isolated a new cysteine derivative, spongiacysteine (1). In this report, we describe the isolation, structural elucidation, and synthesis of spongiacysteine.

Spongia sp. (1.9 kg, wet weight) was extracted with methanol. The methanol extracts were partitioned between H<sub>2</sub>O and EtOAc, and the EtOAc extracts were partitioned between 90% aq methanol and hexane. The 90% aq methanol layers were concentrated and separated by a series of chromatographic processes, including column chromatography (SiO<sub>2</sub> and ODS) and HPLC on ODS to give spongiacysteine (1) (2.0 mg).



Spongiacysteine (1) was isolated as a colorless oil. Its molecular formula was determined to be  $C_{17}H_{33}NO_5S$  based on HRFABMS  $[m/z 386.1972 (M + Na)^+, \Delta -0.5 \text{ mmu}].^3$  The IR spectrum showed the presence of hydroxy (3500–3300 cm<sup>-1</sup>) and carbonyl (1680 and 1650 cm<sup>-1</sup>) moieties. Its <sup>1</sup>H and <sup>13</sup>C NMR spectral data are summarized in Table 1.



**Figure 1.** Partial structures of spongiacysteine (1) based on  ${}^{1}\text{H}{-}^{1}\text{H}$  COSY (bold line) with important HMBC correlations (arrows).

The signals in the <sup>1</sup>H and <sup>13</sup>C NMR spectra were completely assigned by the analysis of the <sup>1</sup>H–<sup>1</sup>H COSY, HMQC, and HMBC spectral data to suggest a planar carbon framework for spongiacysteine (1) as a novel cysteine derivative, as shown in Figure 1. For determination of the absolute stereochemistry, spongiacysteine (1) was converted into the MTPA esters (Figure 2). Chemical shift differences between the two esters gave conflicting results due to the double esterification of the hydroxy functions at C-2 and C-5. However, these data could be explained by separately considering the influence of each MTPA group on the adjacent hydrogens and referring to the examples<sup>4–6</sup> of the modified Mosher's method on a 1,4-diol. The absolute stereochemistry of the two hydroxy moieties was assigned as 2*S*, 5*R*. Subsequently, NMR analysis of the seven-membered lactone **2** derived from **1** (2,4,6-trichlorobenzoyl chloride, DMAP, Et<sub>3</sub>N)<sup>7,8</sup> revealed that the relative stereochemistry between C-2 and C-3 was determined to be anti stereochemistry, establishing that the absolute stereochemistry of C-3 was 3*S* (Figure 3).

Table 1. NMR spectral data of spongiacysteine (1) in CD<sub>3</sub>OD

С	$^{13}C^a$	<sup>1</sup> H <sup>b</sup> / ppm	HMBC
No.	/ ppm	(mult., $J/Hz$ )	$^{13}\mathrm{C} \rightarrow \ ^{1}\mathrm{H}$
1	19.4	1.22 d (6.3)	
2	71.6	3.91 dq (4.1, 6.3)	H-1
3	51.1	2.91 ddd (10.9, 4.1, 3.0)	H-1, 4, 3'
4	39.2	1.67 ddd (14.5, 10.9, 3.0)	H-2
		1.39 m	
5	69.7	3.83 m	H-4, 6
6	41.7	1.45 m	H-4, 7, 8
		1.38 m	
7	20.5	1.46 m	H-6, 8
		1.37 m	
8	14.9	0.92 t (6.9)	H-6, 7
1'	173.4		H-2', 3'
2'	59.2	5.11 dd (11.1, 4.5)	H-3', N-CH <sub>3</sub>
3'	32.2	3.25 dd (14.0, 4.5)	H-2'
		3.00 dd (14.0, 11.1)	
1″	176.7		H-2', 2", 3", N-CH <sub>3</sub>
2"	43.7	2.34 dd (14.9, 7.3)	H-3", 4", 5"
		2.30 dd (14.9, 6.8)	
3″	27.3	2.13 m	H-2", 4", 5"
4", 5"	23.4	1.00 d (6.6)	H-2", 3", 4", 5"
N-CH <sub>3</sub>	34.2	3.04 s	H-2′

<sup>a</sup> Recorded at 125 MHz, <sup>b</sup> Recorded at 500 MHz.



**Figure 2.** Chemical shift differences  $(\Delta \delta)$  between the MTPA derivatives of spongiacysteine (1) [500 MHz, CDCl<sub>3</sub>].

We could not obtain any information of the stereochemistry of C-2', however, we can postulate that the absolute stereochemistry of the cysteine moiety would be L as in the case of acyclic natural products. To confirm the absolute stereostructure, we decided to synthesize 1 stereoselectively (Scheme 1).

*N*-Methylcysteine  $(3)^9$  reacted with isovaleroyl chloride to



Figure 3. Selected NOESY correlations and coupling constants of the lactone 2.

give the amide 4. Kinetic resolution of the racemic 1,2-epoxypentane  $[(\pm)-5]$  in the presence of the asymmetric catalyst 6 provided the optically active epoxide (+)-5.<sup>10</sup> The coupling reaction of the epoxide (+)-5 with lithium acetylide followed by methylation provided the alcohol 7. Hydrogenation of the alcohol 7 in the presence of the Lindlar catalyst led to a homoallylic alcohol, the diastereoselective epoxidation of which provided the epoxide 8 in 99% de.11

Coupling of the amide 4 and the epoxide 8 was accomplished and produced spongiacysteine  $(1)^{12}$  along with the regioisomer 9. The <sup>1</sup>H NMR spectral data of the synthetic spongiacysteine was identical to those of the natural compound. Furthermore, to confirm the stereochemistry in such an acyclic system, synthetic 1 was converted into a seven-membered lactone 2, whose spectral data were found to be identical with those of the authentic sample from natural 1.



Scheme 1. Reagents and conditions: (a) isovaleroyl chloride, DMAP, pyridine, rt, 30 min, 23%; (b) (*R*,*R*)-6, H<sub>2</sub>O, 0 °C  $\rightarrow$ rt, 22h, 35%, >99% ee; (c) lithium acetylide EDA complex, DMSO, rt, 2h, 99%; (d) methyl iodide, n-BuLi, THF, -78°C  $\rightarrow$  rt, 4 h, 92%; (e) H<sub>2</sub>, Lindlar catalyst, MeOH, rt, 6 h, 83%; (f) t-BuOOH, VO(acac)<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 3 h, 99%, >99% de; (g) 1 M NaOH, THF-H<sub>2</sub>O (1:1), 50 °C, 16 h, **1** (41%), **9** (36%).

In conclusion, we isolated a novel cysteine derivative, spongiacysteine (1), from marine sponge Spongia sp. The gross structure was elucidated by detailed spectroscopic analyses, and the absolute stereostructure was established by its total synthesis. As a result of an investigation of various biological activities of 1, it turns out that 1 showed antimicrobial activity against rice blast fungus *Pyricularia oryzae* at  $IC_{90} = 100$  ppm. Blasticidin, kasugamycin, etc.,<sup>13</sup> are known to possess the same activity and used in crop field. Spongiacysteine (1) has a different type of structure from them and was expected to be a lead for new types of agricultural chemicals. In our laboratory, we plan to synthesize its analogues and investigate their biological activities.

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## **References and Notes**

- H. Kigoshi, H. Niwa, K. Yamada, T. J. Stout, and J. Clardy, Tetrahedron Lett., 32, 2427 (1991).
- 2 This sponge was identified as a new species in Spongia by
- Professor Patricia R. Bergquist, the University of Auckland. Spongiacysteine 1:  $[α]^{20}_D 238$  (*c* 0.02, MeOH); FT/IR (neat)  $ν_{max}$  3500–3300 (br.), 2928, 1717, 1684, 1652, 1616, 1135 cm<sup>-1</sup>. 3
- I. Ohtani, T. Kusumi, Y. Kashman, and H. Kakisawa, J. Am. Chem. 4 Soc., 113, 4092 (1991).
- A. Fontana, M. C. Gonzalez, M. Gavagnin, J. Templado, and G. 5 Cimino, Tetrahedron Lett., 41, 429 (2000).
- In the modified Mosher's method, the effects of a MTPA ester were thought to appear more strongly at the  $\gamma$ - and  $\mathcal{E}$ -hydrogens than at the  $\beta$ - and  $\delta$ -hydrogens in acyclic compounds. In spongiacysteine (1), we could conclude that H-4 was strongly influenced by the MTPA ester at C-2. On the other hand, H-3 was influenced by the MTPA ester at C-5. The positional differences of the effect of the MTPA group are found in many literatures. See: T. Kusumi, T. Ooi, and H. Uchimura, Tetrahedron Lett., 35, 3127 (1994); T. Kusumi, H. Takahashi, P. Xu, T. Fukushima, Y. Asakawa, T. Hashimoto, Y. Kan, and Y. Inouye, Tetrahedron Lett., 35, 4397 (1994); K. Kouda, T. Ooi, and T. Kusumi, Tetrahedron Lett., 40, 3005 (1999).
- 7 a) M. Hikota, H. Tone, K. Horita, and O. Yonemitsu, J. Org. Chem., 55, 7 (1990). b) J. Inanaga, K. Hirata, H. Saeki, T. Katsuki, and M. Yamaguchi, Bull. Chem. Soc. Jpn., 52, 1989 (1979).
- **2**: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  5.79 (dd, J = 2.6, 7.3 Hz, H-2', 1H), 4.69 (dq, J = 9.8, 6.3 Hz, H-2, 1H), 3.90 (m, H-5, 1H), 3.26 (dd, J = 7.3, 15.3 Hz, H-3' $\alpha$ , 1H), 3.17 (ddd, J = 2.4, 9.8, 13.8 Hz, H-3, 1H), 3.12 (s,  $N-CH_3$ , 3H), 2.72 (dd, J = 2.6, 15.3 Hz, H-3' $\beta$ , 1H), 2.28 (dd, J = 6.9, 14.8 Hz, H-2" $\alpha$ , 1H), 2.22 (dd, J = 6.8, 14.8 Hz, H-2" $\beta$ , 1H), 2.17 (m, H-3", 1H), 1.65-1.20 (m, H-4, H-6, H-7, 6H), 1.45 (d, J = 6.3 Hz, H-1, 3H), 0.98 (d, J = 6.4 Hz, H-4", 3H), 0.97 (d, J = 6.4 Hz, H-5", 3H), 0.93 (t, J = 7.1 Hz, H-8, 3H).
- W. Keller-Schierlein, M. L. Mihailovie, and V. Prelog, Helv. Chim. Acta, 26, 305 (1959).
- 10 a) M. Tokunaga, J. F. Larrow, F. Kakiuchi, and E. N. Jacobsen, Science, 227, 936 (1997). b) S. E. Schaus, B. D. Brandes, J. F. Larrow, M. Tokunaga, K. B. Hansen, A. E. Gould, M. E. Furrow, and E. N. Jacobsen, J. Am. Chem. Soc., 124, 1307 (2002).
- 11 E. D. Mihelich, K. Daniels, and D. J. Eickhoff, J. Am. Chem. Soc., 103, 7690 (1981).
- 12 Synthetic spongiacysteine:  $[\alpha]^{24}_{D}$  –115 (*c* 0.02, MeOH). The difference in the values between the natural and synthetic samples may be due to an error in the measurement of the natural specimen due to lack of the sample.
- T. Ishiyama, I. Hara, M. Matsuoka, K. Sato, S. Shimada, R. Izawa, 13 T. Hashimoto, M. Hamada, Y. Okami, T. Takeuchi, and H. Umezawa, J. Antibiot., Ser. A, 18, 115 (1965).